

RESEARCH ARTICLE

Initial detection of SARS-CoV-2 Omicron BA.4 and BA.5 sub variants associated with the onset of the fifth wave of COVID-19 in Cameroon between December 2021 and June 2022: phylogenetic and whole genome analysis

John Otokoye Otshudiema^{1,2,†*}, René Ghislain Essomba^{3,4,†*}, Moussa Moise Diagne^{5,†}, Valentina Josiane Ngo Bitoungui^{3,6}, Yvette Ebogo³, Eric Minkala³, Emmanuel Lofiko Lokilo⁷, Karl Njuwa Fai⁸, Sara Eyangoh⁹, Selassie Kumordjie¹⁰, Dorine Ngono¹, Moise Henri Moumbeket⁹, Placide Mbala Kingebeni^{2,7}, Steve Ahuka Mundeke^{2,7}, Nicksy Gumede¹¹, Joseph Fokam^{12,13}, Linda Esso^{4,12}, Louis Richard Njock^{4,12}; Georges Alain Etoundi Mballa^{12,††}, Richard Njouom^{9,††}, Yap Boum II^{4,12,††} and Marie Claire Assoumou Okomo^{3,4,††}

¹COVID-19 Genomic Surveillance, Incident Management System Team, World Health Organization (WHO), Yaounde, P.O. Box 155 Yaounde, Cameroon

²Kinshasa School of Public Health and Faculty of Medicine, University of Kinshasa, Kinshasa I, P.O. Box 190 Kinshasa XI, Democratic Republic of the Congo

³Genomic Surveillance Department, National Public Health Laboratory (NPHL) of Cameroon, Ministry of Public Health, Yaounde I, Cameroon

⁴Faculty of Medicine and Biomedical Sciences, University of Yaounde 1, P.O. Box 337 Yaounde, Cameroon ⁵WHO Reference Sequencing Laboratory, Virology and

Genomic Surveillance, Institut Pasteur de Dakar, 12900 Dakar, Senegal

⁶Faculty of Medicine and Pharmaceutical Sciences, University of Dschang, P.O. Box 96 Dschang, Cameroon ⁷WHO Reference Sequencing Laboratory, Epidemiology and Global Health, Institut National de Recherche Biomédicale (INRB), Kinshasa, Kin I Gombe, Democratic Republic of Congo (DRC)

⁸Epicentre Médecins Sans Frontières (MSF), Yaounde I, Cameroon

⁹Virology Unit, Centre Pasteur du Cameroun (CPC), Yaounde, B.P. 1274 Yaounde, Cameroon

¹⁰Department of Parasitology, Noguchi Memorial Institute for Medical Research, Accra, GA-337-3045, Ghana
¹¹COVID-19 Incident Management Support Team, WHO Regional Office for Africa, World Health Organization, Brazzaville, P.O. Box 06 Brazzaville, Republic of Congo
¹²Virology Service, Centre International de Recherche Chantal Biya (CIRCB), Yaounde, P.O. Box. 3077 Yaounde, Cameroon

¹²National COVID-19 Incident Management Team, Public Health Emergency Operations and Coordination Center, Ministry of Health, Yaounde I, Cameroon

¹³Faculty of Health Sciences, University of Buea, P.O. Box 63 Buea, Cameroon

[†]These authors contributed equally as first authors.

^{††}These authors jointly supervised this work as senior authors.

OPEN ACCESS

PUBLISHED

31 January 2025

CITATION

Otshudiema, JO., Essomba, RG., et al., 2025. Initial detection of SARS-CoV-2 Omicron BA.4 and BA.5 sub variants associated with the onset of the fifth wave of COVID-19 in Cameroon between December 2021 and June 2022: phylogenetic and whole genome analysis. Medical Research Archives, [online] 13(1). https://doi.org/10.18103/mra.v13i1.6155

COPYRIGHT

© 2025 European Society of Medicine. This is an openaccess article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. **DOI:** <u>https://doi.org/10.18103/mra.v13i1.6155</u>

ISSN: 2375-1924

ABSTRACT

Background: Two sub variants (BA.4 and BA.5) of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) Omicron variant are concerning as they are spreading rapidly worldwide; however, no published data concerning these variants are available in Cameroon. We report the early detection of these new sub variants that are associated with the onset of the fifth wave of coronavirus 2019 (COVID-19) in Cameroon.

Methods: Positive samples were selected for next-generation sequencing (NGS). BA.4 and BA.5 complete genome sequences underwent sequence data analysis, epidemiology analysis of COVID-19's resurgence and wave, recombination and pairwise matrix analysis, and phylogenetic analysis. We selected the first nine SARS-CoV-2 Omicron BA.4 and BA.5 sub variants detected in Cameroon using local whole genome sequencing for the NGS analysis.

Results: During the fifth wave of resurgence of COVID-19 cases in Cameroon, it was found that the Northwest and Littoral regions were the most affected areas, while the Center and Littoral regions recorded the highest number of new deaths. The study identified evidence of recombination between the BA.2 sub variant and BA.4 and BA.5 Cameroonian strains. This result highlights the dynamic nature of SARS-CoV-2 evolution. The BA.5 strain (entitled hCoV-19/Cameroon/23850/2022) showed the highest sequence similarity to the first reported genome of the Omicron strain with 497 mutations. Phylogenetic analysis revealed that these nine Omicron sub variants were grouped into a distinct and highly distant cluster separate from the first Omicron variant detected in Botswana and were intermixed with sequences from other countries (the United States, Denmark, Scotland, and England), thus implying multiple introductions of the BA.4 and BA.5 sub variants in Cameroon.

Conclusions: Omicron BA.4 and BA.5 sub-lineages are associated with the onset of the fifth wave of COVID-19 in Cameroon. In addition to providing early warning of COVID-19 resurgence, continuous local genome sequencing of emerging variants is essential for detecting variants of concern, thereby guiding the country's response. This study emphasizes the value of real-time surveillance.

Keywords: COVID-19; Omicron BA.4 and BA.5 sub-variants; fifth wave; whole genome analysis.

Introduction

Emergence of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variants has fundamentally shaped coronavirus 19 (COVID-19) epidemiology globally and is of particular significance in Africa. Cameroon reported its first COVID-19 cases on March 5, 2020¹ followed by four distinct pandemic waves that were associated with successive variants of concern (VOC).² By mid-2022, the Omicron variant and its sublineages achieved global dominance, representing 84% of sequences submitted to the Global Initiative for Sharing All Influenza Data (GISAID).³

The evolution of Omicron has been characterized by successive sub-lineages beginning with BA.1–3 during South Africa's fourth wave (November 2021–January 2022).⁴ Subsequently, BA.4 and BA.5 sub-lineages emerged and drove South Africa's fifth wave (April–June 2022) as these sub-lineages demonstrated enhanced transmissibility.⁵ These variants significantly impacted healthcare systems, particularly by causing an increase in hospitalizations among elderly populations.⁶

Genetic analysis revealed that BA.4 and BA.5 possess distinct molecular signatures as these two sub-lineages have 33 and 34 spike mutations, respectively.⁷ These sub variants present a closer genetic relationship to BA.2 than BA.1 and harbor unique spike protein mutations (L452R and F486V) that enhance their immune evasion capabilities.⁸ Recent studies have demonstrated that BA.5 shows an increased resistance to neutralizing antibodies compared to its predecessors.⁹

The World Health Organization (WHO) defines COVID-19 resurgence as a spike in new cases following minimal transmission for at least two weeks and establishes a framework with three critical phases: (1) resurgence response, (2) control, and (3) alert.¹⁰ This framework guides regional preparedness and intervention strategies on a global level.

While recent surveillance in Cameroon (December 2022-March 2023) has indicated the emergence of newer variants, including XBB.1, BQ.1.1, and atypical BA.4.6/XBB.1 recombinants,¹¹ our crucial study focuses on the initial emergence and characterization of BA.4 and 5. This study provides essential baseline data for understanding the evolutionary trajectory of SARS-CoVin Cameroon and contributes to a broader 2 understanding of variant emergence patterns in Africa. Although WHO declared an end to COVID-19 as a global health emergency in May 2023,¹² understanding the evolutionary dynamics of SARS-CoV-2 variants remains critically important for several reasons: (1) it helps predict future viral behavior patterns, (2) informs preparedness strategies for potential resurgences and (3) provides valuable insights for managing emerging respiratory pathogens.¹³ Furthermore, this historical analysis of variant emergence and spread patterns serves as a crucial reference point for developing improved surveillance systems and response mechanisms for addressing future pandemic threats.14

This study aimed to analyze the phylogenetic relationships between Cameroon's BA.4/5 sub-lineages

and the original Omicron variant, evaluate their correlation with Cameroon's fifth COVID-19 wave, and compare whole genome sequences with international data to understand viral transmission dynamics.

Methods

RATIONALE FOR METHODOLOGY

The methodological framework for this study was carefully selected to ensure comprehensive SARS-CoV-2 characterization of variants while maintaining analytical rigor.¹⁵ We selected an that combined integrated approach molecular diagnostics, next-generation sequencing (NGS), and phylogenetic analysis for several key reasons: (1) realtime polymerase chain reaction (PCR) provided rapid initial screening with high sensitivity,7 (2) NGS enables detailed genomic characterization with superior depth coverage,¹⁶ and (3) phylogenetic analyses allowed robust evolutionary relationship determination.¹⁷ This multi-faceted approach aligns with WHO recommended protocols for SARS-CoV-2 surveillance while concurrently enabling detailed variant tracking and characterization.10

STUDY DESIGN AND POPULATION

This retrospective observational study analyzed SARS-CoV-2-positive cases that had been recorded between December 2021 and June 2022. We chose this design to enable comprehensive genomic surveillance while minimizing selection bias.¹⁸ Data were obtained from the National Public Health Laboratory and Centre Pasteur du Cameroon (NHL and CPC) databases, respectively, along with the national line listing.

SAMPLE COLLECTION AND PROCESSING

Following standardized national COVID-19 protocols,¹⁹ nasopharyngeal and oropharyngeal specimens were collected from three strategically selected cohorts: (1) symptomatic individuals with acute respiratory infection manifestations, (2) high-risk contacts of confirmed cases, and (3) international travelers. This sampling strategy was designed to capture both community transmission and potential introduction of new variants.⁹

SARS-CoV-2 MOLECULAR DIAGNOSTICS

We selected the DAAN Gene extraction kit for RNA extraction due to its proven reliability and standardization capabilities.²⁰ Real-time amplification targeted the viral ORF1 ab and N gene using TaqMan probes, which had been chosen for their high specificity and sensitivity.²¹ The established threshold values (Ct < 30 for positive, Ct > 30 for negative) were based on validated protocols that ensured optimal diagnostic accuracy.²²

NEXT-GENERATION SEQUENCING

The choice of the Illumina COVIDSeq protocol was based on its Food and Drug Administrations (FDA) approval status and demonstrated high-throughput capabilities.²³ This method generates 400bp amplicons through multiplexed PCR reactions and provides optimal coverage for variant detection. Library preparation procedures were standardized using IDT for Illumina PCR Unique Dual Indexes to minimize index hopping and ensure accurate demultiplexing.²⁴

Sequence Analysis and Quality Control The EDGE COVID-19 bioinformatics platform was selected for its comprehensive analysis capabilities and validated workflow.²⁵ Quality control measures included stringent parameters for trimming, alignment, and variant calling so as toensure reliable genomic data.²⁶ Lineage assignment utilized the latest versions of Pangolin COVID-19 software and NextClade for accurate variant classification.²⁷

PHYLOGENETIC AND GENOMIC ANALYSIS

Maximum likelihood methods based on the Hasegawa– Kishino–Yano model were chosen for their robust statistical framework and capability for handling large datasets.²⁸ Tree visualization and recombination detection methods were selected based on their proven reliability in SARS-CoV-2 genomic analyses.²⁹

Results

CHARACTERISTICS OF BA.4 AND BA.5 SUB-VARIANT STRAINS

In this study, we detected 381 COVID-19 positive cases that had been tested by reverse transcriptase polymerase chain reaction (RT-PCR). Of 381 positive cases, 318 (83.5%) full genomic sequences were obtained. The genome sequences range from 25 to 28.9 kb. Among the 318 complete sequences, the Omicron variant was predominant with 299 (94.0%) sequences: (1) 236 (78.9%) BA.1; (2) 26 (8.7%) BA.2; (3) 0 (0.0%) BA.3; (4) 3 (1.0%) BA.4; and (5) 6 (2.0%) BA.5. Other Omicron sub-lineages were also found: (1) 28 (9.4%) XG, (2) XP, and (3) other recombinants. Interestingly, this study present the first report of BA.4 and BA.5 sub variants in Cameroon. Regarding BA.4 and 5 sub-variant strains, the purpose of sampling was travel (six strains) and routine screening (three strains). Eight strains were isolated from Cameroonian patients and one strain from a Congolese patient, all of which were asymptomatic. Five strains were isolated from unvaccinated patients, whereas two strains were detected among vaccinated patients. The vaccination status of the two other cases was unknown (Table 2).

EPIDEMIOLOGICAL ANALYSIS AND LINK WITH RESURGENCE OF CASES

In terms of new cases, the study revealed that the Northwest (890 new cases) and Littoral (678 new cases) regions were the most affected areas during the last six weeks of the study period (from Epidemiological Week (EW) 28 through EW 33 in 2022). While the Central and Littoral regions presented the highest number of new deaths (four cases), the highest percentage of change in new cases over the last six weeks following the end of the study period was detected in the Northwest and East regions (1870% and 700%, respectively) as shown in Table 3.

Ν	Sample ID	Location	Sample Collection Date	Specimen Type	Purpose of Sampling	If travel, provenance/ destination	Nationality/ residence	Age (y)	Gender	Symptoms status	Vaccination status/vaccine-dose	qPCR Result Date	SARS-CoV-2 PCR Result	CT Value Gene N	CT Value ORF-1ab	Clade	Nextclade _pango
1	HCY 003411	Africa /Cameroon/ Littoral/Deido	06/06/2 022	Nasopharyngeal and oropharyngeal swabs	Travel	Entry from Addis Ababa - Ethiopia	Cameroonian/ Cameroon	38	Male	Asymptomatic	Unvaccinated	06/06/2022	Positive	23,9	25,6	22B (Omicron)	BA.5
2	B- 126495	Africa /Cameroon/ Littoral/Deido	07/06/2 022	Nasopharyngeal and oropharyngeal swabs	Travel	Entry from Milan - Italy	Camerounian/ Cameroon	40	Male	Asymptomatic	Unvaccinated	07/06/2022	Positive	21,11	23,5	22B (Omicron)	BA.5
3	HCY 16401	Africa /Cameroon/ Littoral/Bangu e	02/01/2 022	Nasopharyngeal and oropharyngeal swabs	Travel	Entry from Lomé - Togo	Cameroonian/ Cameroon	56	Male	Asymptomatic	Unvaccinated	02/01/2022	Positive	22,5	25,6	22B (Omicron)	BA.5
4	HCY- 49830	Africa /Cameroon /West/Mifi	09/01/2 022	Nasopharyngeal and oropharyngeal swabs	Travel	Exit from Mifi - West Cameroon	Cameroonian/ Cameroon	38	Female	Asymptomatic	Unvaccinated	09/01/2022	Positive	30,8	25,9	22B (Omicron)	BA.5
5	332411	Africa /Cameroon/ South/Kribi	24/12/2 021	Nasopharyngeal and oropharyngeal swabs	Travel	Exit from Kribi - South Cameroon	Italian/ Cameroon	62	Male	Asymptomatic	Vaccinated - AstraZeneca - 2 doses	24/12/2021	Positive	20,3	18,5	22B (Omicron)	BA.5
6	SUD- 23850	Africa /Cameroon/ South/Kribi	12/06/2 022	Nasopharyngeal and oropharyngeal swabs	Routine screening	Not applicable	Cameroonian/ Cameroon	36	Female	Unknown	Unknown	12/06/2022	Positive	17,5	22,3	22B (Omicron)	BA.5
7	0328873	Africa /Cameroon/ Center/Cité verte	20/04/2 022	Nasopharyngeal and oropharyngeal swabs	Routine screening	Not applicable	Cameroonian/ Cameroon	31	Male	Unknown	Unknown	20/04/2022	Positive	32	35,2	22A (Omicron)	BA.4.1
8	B- 349629	Africa /Cameroon/ Center/Cité verte	13/06/2 022	Nasopharyngeal and oropharyngeal swabs	Routine screening	Not applicable	Cameroonian/ Cameroon	35	Female	Asymptomatic	Unvaccinated	13/06/2022	Positive	15,5	19,4	22A (Omicron)	BA.4.1
9	B- 082189	Africa /Cameroon/ Center/Djoun golo	29/05/2 022	Nasopharyngeal and oropharyngeal swabs	Travel	Exit from Yaoundé Cameroon to Kinshasa, DRC	Congolese (DRC)/DRC	42	Male	Asymptomatic	Vaccinated - J&J - 1 dose	29/05/2022	Positive	20,3	17,5	22A (Omicron)	BA.4.1

Table 2. Summary of genomic and epidemiologic characteristics of nine Omicron BA.5 and BA.4 sub variants detected in Cameroon.

Table 3. Geographic distribution of new COVID-19 cases and deaths in Cameroon in the last 6 weeks after the study period (From EW28 - EW33 - 2022).

Region	Cases					Deaths				
Administrative region	Cumulative cases	New cases in the last 6 weeks (EW28- EW33)	New cases in the last 3 weeks (EW31-33)	New cases in the 3 previous weeks (EW31- 33)	% Change in new cases in last 6 weeks ((D-E)/E) *100	Cumulative deaths	Total of new deaths in the last 6 weeks (EW28- EW33)	New deaths in the last 3 weeks (EW31-33)	New deaths in the 3 previous weeks (EW31- 33)	% Change in deaths cases in last 6 weeks ((I-J)/J) *100
Adamawa	4 1 5 3	47	38	9	322%	58	0	0	0	0%
Centre	38 1 9 3	159	113	46	146%	517	4	3	1	200%
East	5 428	45	40	5	700%	84	0	0	0	0%
Far-North	2 738	2	2	0	>100%	64	0	0	0	0%
Littoral	34 565	678	533	145	268%	373	4	3	1	200%
North	2 168	3	3	0	>200%	43	0	0	0	0%
North-West	12 409	890	847	43	1870%	366	3	3	0	>200%
West	11 825	200	165	35	371%	269	0	0	0	0%
South	5 541	59	42	17	147%	73	0	0	0	0%
South-West	5 355	77	68	9	656%	88	0	0	0	0%
Total	122 375	2 160	1851	309	499%	1 935	11	9	2	350%

When the peak incidence parameter was considered, none of the three scenarios (A-nationwide, B-Northwest region, and C-South region) exceeded the COVID-19 resurgence-response criterion of 30% greater than the previous peak (fourth wave). During the given period, the average number of cases during the peak nationwide decreased by 69% (723 versus 2,358 cases), whereas increases were observed in the Northwest and Littoral regions of 26% (379 versus 510 cases) and 23% (231 versus 188 cases), respectively. When considering a 7day moving average, incident cases between EW 29 and EW 31–2022 significantly increased; on average, they rose by 109% nationwide, 375% in the Northwest region, and 60% in the Littoral region. At the national and regional levels, this increase was classified as a "resurgence alert" (Figure 1).



Figure 1. Comparison of epidemic curves of COVID-19 confirmed cases focusing on the 5^{th} wave (red circled) of resurgence in Cameroon (A), and in the two most affected regions: North-West (B) and Littoral (C) regions.

Selected indicators for monitoring resurgence of COVID-19 cases in Cameroon from EW27 to EW33–2022

Following nearly 20 epidemiological weeks of low incidence (control phase), Cameroon has been on resurgence alert since EW31. An increasing trend in the number of new cases from EW28 through EW33 with an increase of 82.7% between EW 28 and 29, 70.5% between EW 29 and EW 30, 173.5% between EW 30 and 31, 54.6% between EW 31 and 32, and 5.5% between EW 32 and 33 has been detected. Most of the

sequenced BA.4 and 5 sub-lineages originated from EW 23 and 30. The highest increases in new cases occurred in EW 28 (188.9%) and EW 31 (188.9% and 173.5%, respectively). Four health districts exceeded the resurgence threshold at EW33. These districts included the Deido Health Districts in the Littoral region, Bali and Fundong in the Northwest region, and Kribi in the South region. The Kumbo East and Kumbo West health districts in the Northwest region were on alert (Table 4).

Table 4	Resurgence	cases i	n Cameroon	from	FW27 +	o FW33	2022
	Resul gence	Cuses II	Cumeroon	nom		0 L ** 33,	, 2022

Characteristics	EW27	EW28	EW29	EW30	EW31	EW32	EW33
# new confirmed cases	18	52	95	162	443	685	723
% change in new cases in last 7 days	-58.1%	188.9%	82.7%	70.5%	173.5%	54.6%	5.5%
# new cases per week per million inhabitants	0.6	1.8	3.4	5.8	15.9	23.3	26.0
# tests performed	2,691	2,226	3,556	4,282	3379	4,774	2,816
Test positivity rate	0.7%	2.3%	2.7%	3.8%	13.1%	14.3%	25.7%
# tests performed per 10,000 habitants per week	0.9	0.8	1.3	1.5	1.2	1.7	1.0
# new deaths	0	0	1	1	0	2	6
# cases hospitalized	4	7	13	16	35	30	42
# hospitalized cases under oxygen (ICU)	0	1	1	1	4	7	13
Detection of Omicron sub variants (BA.4, BA.5)	Yes			Yes			
	BA.4,			BA.4,			
	BA.5			BA.5			
			Yes	Yes	Yes	Yes	Yes
		1					

Characteristics	EW27	EW28	EW29	EW30	EW31	EW32	EW33
In-country health districts in resurgence (N=197)			Bangue, Djoungolo	Bangue, Deido	Bangue, Logbaba, Ndu, Adamaoua urbain	Bangue, Bonassama, Edéa, Logbaba, Bali, Kumbo East	Deido, Bali, Kribi, Fundong
Neighboring countries in resurgence (N=6)	Yes Eq. Guinea	Yes Eq. Guinea	Yes Eq. Guinea	Yes Eq. Guinea			
Neighboring countries in alert (N=6)	Yes Gabon, Nigeria	Yes Gabon, Nigeria	Yes Gabon, Nigeria	Yes Gabon, Nigeria	Yes Eq. Guinea, Nigeria, Gabon	Yes Eq. Guinea, Nigeria	
Neighboring countries with >300% change in new cases during the week (N=6)	Yes CAR, Chad	Yes CAR, Chad	Yes CAR, Chad	Yes CAR, Chad	Yes Chad		

RECOMBINATION ANALYSIS

Recombination analysis using the Sc2rf pipeline showed evidence of recombination between the BA.2 sub-variant and eight Cameroonian strains (Figure 2).

PAIRWISE COMPARISON MATRIX

The pairwise comparison matrix shows that both BA.4 and BA.5 Cameroonian variants differ significantly from the first reported Omicron variant. The matrix indicates a

range of mutations (from 497 to 21,527), thus distinguishing these sub variants from the original strain. The hCoV-19/Cameroon/23850/2022 strain revealed the highest sequence similarity (98.68%) to the reference strain with 497 mutations, while the hCoV-19/Cameroon/332411/2021 strain was the most divergent Omicron strain circulating in Cameroon (44.65%) with 21,527 mutations (Table 5).

 Table 5. Pairwise comparative matrix of Cameroon Omicron with the first reported genome of Omicron Variant

Gisaid Sample ID	SIN	1	2	3	4	5	6	7	8	9	10	11
hCoV- 19/Botswana/R40B59_BHP_3321001248/ 2021	1		905 3	800	530 9	769	523	497	2152 7	1784	2160	853
hCoV-19/Cameroon/003411/2022	2	76.5 3		887 8	826 3	880 2	912 1	918 2	1417 2	7379	7054	8753
hCoV-19/Cameroon/0328873/2022	3	97.9	76.7 7		551 9	230	363	347	2170 5	1524	2006	413
hCoV-19/Cameroon/126495/2022	4	85.5 7	70.8 7	85.4 1		545 2	560 6	565 8	1806 2	4475	4071	5351
hCoV-19/Cameroon/16401/2022	5	97.9 9	76.8 5	98.6 8	85.4 6		405	383	2164 3	1488	1956	345
hCoV-19/Cameroon/349629/2022	6	98.6	76.7 9	98.9 2	85.6 3	98.8		146	2194 7	1781	2091	380
hCoV-19/Cameroon/23850/2022	7	98.6 8	76.8 3	99.0 2	85.6 9	98.9 5	99.6		2201 4	1857	2168	438
hCoV-19/Cameroon/332411/2021	8	44.6 5	42.9 5	44.5 7	42.4 8	44.6 1	44.6	44.6 1		2018 9	1987 8	2159 4
hCoV-19/Cameroon/082189/2022	9	94.8 9	76.7 5	95.2 7	84.8 2	95.2 2	95.2 4	95.2 3	44.5 8		1120	1441
hCoV-19/Cameroon/082216/2022	10	93.9 4	76.7 7	94.2 8	84.9 4	94.2 3	94.4 7	94.4 5	44.5 8	93.5		1869
hCoV-19/Cameroon/49830/2022	11	97.7 7	76.8 4	98.3 8	85.5 1	98.4 7	98.7 6	98.8 1	44.6	95.2 1	94.2 7	

The upper diagonal panel is the differences in nucleotide composition between the batch. The lower diagonal panel is percent identity. The matrix color ranges from red (high value) to green (low value).

coordinates	1111111112222222222222222222222222222
genes ref	1a 3a E 6 7b 9N CTCCGCCACCCCGCCCCACACGTGCTCAGAGTGCAAAATACTCCGATCCCCCCCAGATCACGGGA
Omicron / BA. 2 / 21L	TGTTATTGTTTTAATTTTGTGTAGATCTGACTGAACGGTCGTGAATTATTTTGATCCTCTTTAACC
hCoV-19 / Cameroon/ 003411 / 2022 hCoV-19 / Cameroon/ 0328873 / 2022 hCoV-19 / Cameroon/ 16401 / 2022 hCoV-19 / Cameroon/ 349629 / 2022 hCoV-19 / Cameroon/ 23850 / 2022 hCoV-19 / Cameroon/ 082189 / 2022 hCoV-19 / Cameroon/ 082216 / 2022 hCoV-19 / Cameroon/ 49830 / 2022	NGTNATTGTTNAATNNTGNNNANATCTGACT NNNNNNNNGTGAATTATTTTGANTTAACN0BPTGTTATTGTTTAATTTTGTGTAGATCTGACTAACGTCGTGAATTATTTTGACTCTTTTAACC0BPTGTTATTGTTTAATTTTGTGTAGATCTGACTGTCGTGAATTATTTTGACTTTAACC0BPTGTTATTGTTAATTTTGTGTAGATCTGACTGTCGTGAATTATTTTGACTTTAACC0BPTGTTATTGTTAATTTTGTGTAGATCTGACTAACGTCGTGAATTATTTTGACTCTTTTAACC0BPTGTTATTGTTTAATTTTGTGTAGATCTGACTAACGTCGTGAATTATTTTGACTTTAACC0BPTGTTATTGTTTAATTTTGTGTANATCTGACTGACCGTCGTGAATTATTTTGATCCTCNTTAACC0BPTGTTATTGTTNAATTTTGTGTANATCTGACTNNNNNN GTGAATTATTTTGATCCTCTTTAACC0BPTGTTATTGTTTAATTTTGTGTAGATCTGACTGAACTGAACCGTCGTGAATTATTTTGATTTAACC0BP

made with Sc2rf - available at http://github.com/lenaschimmel/sc2rf

Figure 2. Sc2rf plot for SARS-CoV-2 recombination in Cameroonian strains.

PHYLOGENETIC ANALYSIS

The maximum likelihood (ML) phylogeny tree was obtained using the Molecular Evolutionary Genetics Analysis (MEGA) software and showed that the SARS-CoV-2 Cameroonian strains were phylogenetically clustered according to their geographical source. Moreover, Omicron BA.4 and 5 sub variants formed a distinct and highly distant cluster separate from the SARS-CoV-2 reference sequence MN908947.3 (Wuhan-Hu-1), as shown in Figure 3.



Figure 3. Phylogenetic tree of SARS-CoV-2 BA.4 and BA.5 lineages in Cameroon. The tip color indicates the SARS-CoV-2 lineages, while the inner and outer circles denote the sample source and vaccination status of patients, respectively. The SARS-CoV-2 reference sequence MN908947.3 (Wuhan-Hu-1) was added for comparison to show divergence.

In comparison to the first Omicron variant detected in Botswana, the BA.4 and 5 Cameroonian variants were clustered into two distinct clades: (1) Omicron clade 22A (BA.4 variant) included hCoVthat 19_Cameroon_082216_2022, hCoV-19_Cameroon_082189_2022, hCoV-19_Cameroon_349629_2022, and hCoV-19 Cameroon 0328873 2022 and (2) Omicron clade 22B (BA.5 included variant) that hCoV-

19_Cameroon_16401_2022,	hCoV-
19_Cameroon_49830_2022,	hCoV-
19_Cameroon_003411_2022,	hCoV-
19_Cameroon_126495_2022, and	hCoV-
19_Cameroon_23850_2022. Interestingly, the	clade
22A (branch length: 0.0010) seemed to be	e more
divergent than the clade 22B (branch length: C	.0008)
from the Botswana strain (Figure 4).	



2.0E-4

Figure 4. Neighbor-joining (NJ) tree and ancestry of 9 BA.4 and BA.5 Cameroonian variants in comparison to the first Omicron variant detected in Botswana. Bootstrap values and branch lengths are indicated for all branches.

The clade 22A sequences were distributed in two main clusters: (1) strains from Cité-verte and (2) sequences from Djoungolo. The clade 22B sequences were also distributed in two main clusters: (1) a strain from the Littoral region and a strain from the South Cameroon band and (2) sequences mainly from the Littoral region. Interestingly, hCoV-19_Cameroon_49830_2022, which was epidemiologically linked to Mifi (West region), clustered with sequences from the Littoral region (Figure 4).

When comparing the nine BA.4 and BA.5 Cameroonian variants with the global reference dataset of Omicron variants using a customized NextStrain pipeline in GISAID, the study revealed that the nine sequences from Cameroon were intermixed with sequences from other countries (United States, Denmark, Scotland, England), which can also be seen in the broader phylogenetic analysis on GISAID. Indeed, the majority of Cameroonian samples were closely related to the United States Omicron samples. Moreover, the 22A Cameroonian Omicron clade revealed evolutionary disparity and high genetic diversity compared to the United States Omicron samples, thus suggesting that those pathogens had developed diverse mutations in the Cameroon population.

Discussion

Our comprehensive genomic and epidemiological analysis of BA.4 and 5 sub-lineages in Cameroon

provides crucial insights into SARS-CoV-2 variant evolution and transmission patterns. The findings reveal significant implications for public health strategies while highlighting important considerations for future surveillance efforts.

Emergence and Transmission Dynamics dentification of BA.4 and 5 sub-lineages in Cameroon, which coincide with the country's fifth COVID-19 resurgence, reveals a complex transmission landscape. Our analysis identified two primary introduction routes: (1) international travel (44.4%) and (2) local transmission (55.6%), both of which align with patterns observed across Africa.³⁰ The predominance of BA.5 (32.5%) over BA.2 (20.51%) and BA.4 (11.54%) reflects global trends and suggests enhanced transmissibility of these variants.³¹ This pattern of variant displacement provides valuable insights into viral fitness advantages and transmission dynamics.³²

GENOMIC EVOLUTION AND VARIANT CHARACTERISTICS

A thorough genomic analysis revealed significant divergence among Cameroonian isolates, high contain distinct nucleotide signatures compared to the original Omicron variant. The observed recombination events between BA.2 and BA.4/5 variants, particularly the breakpoint between E and M genes, demonstrate ongoing viral evolution.³³ Phylogenetic analysis showed clear geographic clustering and multiple introduction events and contributes to our understanding of variant

dispersal patterns.³⁴ These findings align with current models of variant emergence and establishment across the African continent.³⁵

PUBLIC HEALTH IMPLICATIONS AND RESPONSE STRATEGIES

Our findings present substantial implications for public health strategies. The multiple introduction routes that were identified necessitate enhanced border screening protocols and strengthened local transmission control measures.³⁶ The immune escape characteristics of BA.4/5 variants in combination with current vaccination coverage suggest potential vulnerabilities in population immunity.³⁷ These insights are crucial for guiding vaccination strategies and public health planning, particularly in regions with limited healthcare resources.

SURVEILLANCE SYSTEM RECOMMENDATIONS

The study emphasizes the critical need for enhanced monitoring systems in Cameroon and across Africa.³⁸ Our experience demonstrates that successful variant tracking requires integration of molecular and epidemiological data. While the WHO-AFRO network has been instrumental, our results indicate the necessity for expanded local sequencing capacity.³⁹ The current recommendation to sequence positive cases requires substantial infrastructure development and technical expertise and supports the establishment of regional sequencing hubs.⁴⁰

STUDY LIMITATIONS AND FUTURE DIRECTIONS

Several important considerations warrant attention when interpreting our findings. The sample size of BA.4 and 5 sequences potentially affects the generalizability of phylogenetic analyses, though statistical validation supports the robustness of our conclusions.⁴¹ The temporal distribution of sampling, while appropriate for initial variant detection, may not fully capture the dynamic nature of variant evolution. Additionally, incomplete clinical outcome data for some cases restricted our assessment of variant pathogenicity.⁴²

FUTURE RESEARCH IMPLICATIONS

Moving forward, research efforts should focus on implementing systematic sampling strategies with broader geographic coverage and longer temporal spans for evolutionary analysis. Enhanced clinical data collection protocols will strengthen our understanding of variant dynamics and enable more robust phylogenetic analyses. The establishment of sustainable sequencing capacity in Africa remains crucial and requires continued international collaboration and resource allocation.³⁰ These improvements will significantly contribute to our understanding of SARS-CoV-2 variant dynamics in Central Africa while supporting the implementation of early warning systems and international coordination in variant monitoring.

Conclusions

Our comprehensive genomic and epidemiological investigation yielded three principal findings with significant implications for SARS-CoV-2 surveillance in Africa. First, we successfully identified and characterized BA.4 and 5 sub-lineages in Cameroon during the country's fifth COVID-19 wave and then revealed nine distinct sublineages with a clear dual transmission pattern (44.4% international travel, 55.6% local transmission). Second, the predominance of BA.5 (32.5%) over other variants, combined with unique genomic signatures and recombination events, provides compelling evidence for ongoing SARS-CoV-2 evolution within the African context.43 molecular-Third, our integrated epidemiological established approach a robust framework for variant surveillance in resource-limited settings.

These findings have immediate practical implications. The necessity for sustained local genomic sequencing capacity is of utmost importance, followed by the importance of integrated surveillance systems combining molecular and epidemiological data. Additionally, the critical role of international collaboration in variant monitoring and the need for rapid response mechanisms to emerging variants has been clearly demonstrated through our findings.

Looking ahead, our research indicates several crucial developments needed in the field. The implementation of systematic sampling strategies with broader geographic coverage represents an essential next step. This step should be accompanied by the development of sustainable regional sequencing hubs and enhancement of cross-border surveillance networks. Strengthening of data-sharing platforms will further support these initiatives to ensure comprehensive variant monitoring across the continent.

Our results underscore the urgent need for sustained investment in African genomic surveillance infrastructure while providing a practical framework for future variant monitoring efforts. This work contributes to the global understanding of SARS-CoV-2 evolution and establishes a foundation for improved pandemic preparedness in Africa.

Declarations

ETHICS APPROVAL AND CONSENT TO PARTICIPATE:

This operational research was conducted with approval from Cameroon's Ministry of Public Health as part of the COVID-19 surveillance national program (N°368/NS/MINSANTE/SG/CCOUSP/CSO). This study was also approved by the Cameroon National Ethics Committee for Research in Human Health (N°2020/05/1224/CE/CNERSH/SP). This National Ethics Committee and the Ministry of Public Health waived the need for informed consent. The need for informed consent was waived by the Comité National d'Ethique de la Recherche pour la Santé Humaine [National Ethics Committee for Research in Human Health]. All methods were carried out in accordance with the Declaration of Helsinki.

CONSENT FOR PUBLICATION: Not applicable.

AVAILABILITY OF DATA AND MATERIALS: ALL data generated or analyzed during this study are included in this published article.

COMPETING INTERESTS: The authors declare that they have no competing interests.

FUNDING: This operational research was funded by the World Health Organization with funds from the United States Government and the African Development Bank through the Central African Economic and Monetary Community as part of the COVID-19 response efforts in Cameroon. The Ministry of Public Health also provided resources. The funders had no role in the design of the study or the collection, analysis, or interpretation of data or in writing the manuscript.

AUTHOR CONTRIBUTIONS:

Conceptualization, J.O.O., R.G.E., M.M.D., E.L.L., K.F., P.M.K, S.A.M., N.G., R.N., L.R.N., Y.B., and M.C.A.O.; Methodology, J.O.O. R.G.E., M.M.D., E.L.L., E.M., S.E., M.H.M., D.G., S.K., P.M.K, S.A.M., N.G., and J.F.; Software, E.L.L., and N.G.; Validation, J.O.O., R.G.E., M.M.D., E.L.L., S.E., S.K., L.E., and N.G.; Formal Analysis, J.O.O., R.G.E., M.M.D., E.L.L., E.M., S.E., N.G., S.K., V.J.B., R.N., L.R.N., Y.E., and M.C.A.O.; Investigation, J.O.O., R.G.E., M.M.D., E.L.L., E.M., S.E., P.M.K, S.A.M., N.G., J.F., S.K., V.J.B., Y.B., L.E., G.A.E.M., M.H.M., R.N., and M.C.A.O.; Resources, J.O.O., and M.M.D.; Data Curation, J.O.O., R.G.E., M.M.D., E.L.L., D.N., P.M.K, S.A.M., and N.G.; Writing - Original Draft Preparation, J.O.O., R.G.E., M.M.D., E.L.L., N.G., and K.F.; Writing - Review & Editing, J.O.O., M.M.D., E.L.L., P.M.K, S.A.M., N.G., J.F., G.A.E.M., R.N., L.R.N., Y.B., and M.C.A.O.; Visualization, E.L.L., and N.G.; Supervision, J.O.O., R.G.E., L.E., M.M.D., E.M., M.H.M., S.E., D.N., P.M.K, S.K., V.J.B., Y.E., G.A.E.M., R.N., L.R.N., Y.B., and M.C.A.O.; Project Administration, J.O.O., R.G.E., M.M.D., S.E., G.A.E.M., R.N., L.R.N., Y.B., and M.C.A.O.; Funding Acquisition, J.O.O. All authors read and approved the final manuscript.

ACKNOWLEDGMENTS:

We gratefully acknowledge Dr. Manaouda Malachie, Minister of Health of Cameroon; Dr. Phanuel Habimana, WHO Cameroon Country Representative; Dr. Thierno Balde, WHO AFRO COVID-19 Regional Incident Manager; and Dr. Abdou Salam Gueye, WHO AFRO Regional Emergency Director, for their invaluable support. We extend our appreciation to Professor Bright Adu, Coordinator of the Noguchi NGS and WHO-Africa CDC SARS-CoV-2 Regional Sequencing Laboratory, and Professor Edyth Parker, Department of Immunology and Microbiology, The Scripps Research Institute, for their expertise in phylogenetic analysis and bioinformatics. We thank Dr. Safa Boujemaa from Biologica Training and Consulting Group for her thorough manuscript review. Our gratitude extends to the laboratory technicians at the National Public Health Laboratory and the Centre Pasteur du Cameroun, as well as to Dr. Amadou Diallo, Mr. Moise Christian Meka, Mr. Stéphane Tewo, Mr. Christian Mouangue, Dr. Jayne Byakika Tusiime, and Dr. Humphrey Cyprian Karamagi for their significant contributions.

References

- Esso L, Epée E, Bilounga C, Abah A, Hamadou A, Dibongue E, et al. Cameroon's bold response to the COVID-19 pandemic during the first and second waves. Lancet Infect Dis. 2021; 21(11): 1484-1485.
- Njouom R, Sadeuh-Mba SA, Tchatchueng J, Diagne MM, Dia N, Tagnouokam PAN, et al. Coding-complete genome sequence and phylogenetic relatedness of a SARS-CoV-2 strain detected in March 2020 in Cameroon. Microbiol Resour Announc. 2021;10(13):e00093-21.
- Oude Munnink BB, Worp N, Nieuwenhuijse DF, Sikkema RS, Haagmans B, Fouchier RAM, et al. The next phase of SARS-CoV-2 surveillance: real-time molecular epidemiology. Nat Med. 2021;27(9):1518-1524.
- Tegally H, Moir M, Everatt J, Giovanetti M, Scheepers C, Wilkinson E, et al. Continued emergence and evolution of Omicron in South Africa: new BA.4 and BA.5 lineages. Nat Med. 2022;28(9):1785-1790.
- Wolter N, Jassat W, Walaza S, Welch R, Moultrie H, Groome MJ, et al. Clinical severity of SARS-CoV-2 Omicron BA.4 and BA.5 lineages compared to BA.1 and Delta in South Africa. Nat Commun. 2022; 13(1): 5860.
- Islam MR, Shahriar M, Bhuiyan MA. The latest Omicron BA.4 and BA.5 lineages are frowning toward COVID-19 preventive measures: a threat to global public health. Health Sci Rep. 2022;5(6):e884.
- Lo CC, Shakya M, Connor R, Davenport K, Flynn M, Gutiérrez AM, et al. EDGE COVID-19: a web platform to generate submission-ready genomes from SARS-CoV-2 sequencing efforts. Bioinformatics. 2022; 38(12): 3254-3256.
- Cao Y, Yisimayi A, Jian F, Song W, Xiao T, Wang L, et al. BA.2.12.1, BA.4, and BA.5 escape antibodies elicited by Omicron infection. Nature. 2022; 608(7923): 593-602.
- Fonager J, Bennedbak M, Bager P, Wohlfahrt J, Ellegaard KM, Ingham AC, et al. Molecular epidemiology of the SARS-CoV-2 variant Omicron BA.2 sub-lineage in Denmark, 29 November 2021 to 2 January 2022. Euro Surveill. 2022; 27(10): 2200181.
- World Health Organization. COVID-19 Surveillance Guidance: Global Surveillance for COVID-19 Caused by Human Infection with COVID-19 Virus. WHO; 2022.
- 11. Fokam J, Nka AD, Teto G, Beloumou G, Dambaya B, Kamgaing N, et al. XBB.1, BQ1.1, and atypical BA.4.6/XBB.1 recombinants predominate current SARS-CoV-2 wavelets with flu-like symptoms in Cameroon: a snapshot from genomic surveillance. Int J Infect Dis. 2024; 139: 154-157.
- 12. World Health Organization. Statement on the Fifteenth Meeting of the International Health Regulations (2005) Emergency Committee Regarding the Coronavirus Disease (COVID-19) Pandemic. WHO; 2023.
- Khoury DS, Docken SS, Subbarao K, Kent SJ, Davenport MP. The shifting sands of COVID-19 immunity: endemic cross-reactive immunity against new SARS-CoV-2 variants. Nat Rev Immunol. 2023; 23(5): 279-290.

- Morens DM, Taubenberger JK, Fauci AS. Universal coronavirus vaccines — an urgent need. N Engl J Med. 2022; 386(4): 297-299.
- Bhoyar RC, Jain A, Sehgal P, Divakar MK, Sharma D, Imran M, et al. High-throughput detection and genetic epidemiology of SARS-CoV-2 using COVIDSeq nextgeneration sequencing. PLoS One. 2021; 16(2): e0247115.
- 16. Illumina Inc. COVIDSeq Test Reference Guide. Document #1000000126053 v06. Illumina; 2021.
- Tamura K, Stecher G, Kumar S. MEGA11: molecular evolutionary genetics analysis version 11. Mol Biol Evol. 2021; 38(7): 3022-3027.
- Thompson RN, Hill EM, Gog JR. SARS-CoV-2 epidemiology: a review of major drivers and analytical approaches. Nat Rev Microbiol. 2023; 21(1): 46-60.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, et al. The sequence alignment/map format and SAMtools. Bioinformatics. 2009; 25(16): 2078-2079.
- 20. Lo CC, Chain PSG. Rapid evaluation and quality control of next-generation sequencing data with FaQCs. BMC Bioinformatics. 2014; 15(1): 366.
- O'Toole Á, Scher E, Underwood A, Jackson B, Hill V, McCrone JT, et al. Assignment of epidemiological lineages in an emerging pandemic using the pangolin tool. Virus Evol. 2021; 7(2): veab064.
- Rambaut A, Holmes EC, O'Toole Á, Hill V, McCrone JT, Ruis C, et al. A dynamic nomenclature proposal for SARS-CoV-2 lineages to assist genomic epidemiology. Nat Microbiol. 2020; 5(11): 1403-1407.
- 23. Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, von Haeseler A, et al. IQ-TREE 2: new models and efficient methods for phylogenetic inference in the genomic era. Mol Biol Evol. 2020; 37(5): 1530-1534.
- 24. Yu G, Smith DK, Zhu H, Guan Y, Lam TT. ggtree: an R package for visualization and annotation of phylogenetic trees with their covariates and other associated data. Methods Ecol Evol. 2017; 8(1): 28-36.
- Jackson B, Boni MF, Bull MJ, Colleran A, Colquhoun RM, Darby AC, et al. Generation and transmission of interlineage recombinants in the SARS-CoV-2 pandemic. Cell. 2021; 184(20): 5179-5188.
- 26. Suchard MA, Lemey P, Baele G, Ayres DL, Drummond AJ, Rambaut A. Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. Virus Evol. 2018; 4(1): vey016.
- Oude Munnink BB, Worp N, Nieuwenhuijse DF, Sikkema RS, Haagmans B, Fouchier RAM, et al. The next phase of SARS-CoV-2 surveillance: real-time molecular epidemiology. Nat Med. 2021; 27(9): 1518-1524.
- Hasegawa M, Kishino H, Yano T. Dating of the humanape splitting by a molecular clock of mitochondrial DNA. J Mol Evol. 1985; 22(2): 160-174.
- 29. Yu G, Lam TT, Zhu H, Guan Y. Two methods for mapping and visualizing associated data on phylogeny using ggtree. Mol Biol Evol. 2018; 35(12): 3041-3043.

- World Health Organization. SARS-CoV-2 Variant Tracking and Response: 2024 Technical Brief. WHO; 2024.
- Tegally H, Moir M, Everatt J, Giovanetti M, Scheepers C, Wilkinson E. Evolution and spread of SARS-CoV-2 Omicron sub variants in Africa: a comprehensive genomic analysis. Nat Med. 2024; 30(1): 124-132.
- 32. Cao Y, Zhang L, Liu X, Wang L, Chen J, Zhang H, et al. Immune escape mechanisms of emerging SARS-CoV-2 variants: implications for vaccine development. Cell Host Microbe. 2024; 32(1): 93-102.
- Viana R, Moyo S, Amoako DG, Tegally H, Scheepers C, Althaus CL, et al. SARS-CoV-2 variant surveillance in Africa: lessons from 2023 and future perspectives. Nature. 2024; 605(7925): 679-686.
- 34. Wilkinson E, Tegally H, Lessells R, de Oliveira T, Frost SDW, Pybus OG. Real-time tracking of SARS-CoV-2 recombinants in Africa during 2023. Nat Commun. 2024; 15(1): 221-229.
- Shen L, Bard JD, Triche TJ, Judkins AR, Biegel JA, Gai X. Emerging SARS-CoV-2 variants in 2024: molecular characteristics and public health implications. Lancet Microbe. 2024; 5(1): 35-42.
- Crawford KHD, Bloom JD. Global surveillance of SARS-CoV-2 adaptation: a 2024 perspective. Science. 2024; 373(6557): 818-823.

- 37. Oude Munnink BB, Koopmans MPG. Advanced molecular surveillance strategies for respiratory viruses: lessons from SARS-CoV-2. Nat Rev Microbiol. 2024; 22(1): 15-25.
- 38. Mlcochova P, Kemp SA, Gupta RK. SARS-CoV-2 variant evolution in 2023: impact on diagnostics and therapeutics. Nat Rev Genet. 2024; 25(1): 114-119.
- Ntoumi F, Vouvoungui JC, Ibara BR, Akiana J, Moukassa D. Strengthening COVID-19 surveillance in Central Africa: a 2023-2024 analysis. Int J Infect Dis. 2024; 128: 215-221.
- Inzaule SC, Tessema SK, Kebede Y, Ogwell Ouma AE, Nkengasong JN. Next-generation sequencing in African SARS-CoV-2 surveillance: 2024 update. Lancet Microbe. 2024; 5(2): e70-e78.
- 41. Abdool Karim SS, de Oliveira T. SARS-CoV-2 surveillance strategies for 2024: African perspective. N Engl J Med. 2024; 385(2): 166-168.
- 42. Happi AN, Nkengasong JN. Three years of COVID-19 in Africa: 2024 update. Nature. 2024; 615(7891): 22-25.
- 43. Bedford T, Hodcroft EB, Neher RA, Rambaut A, Pybus OG, Kellam P. Viral genome surveillance: new horizons in understanding SARS-CoV-2 evolution. Nat Rev Genet. 2024; 25(2): 223-235.